

# Regional carbon dioxide monitoring to assess the adequacy of tissue perfusion

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## Purpose of review

Tissue dysoxia is now widely regarded as the major factor leading to organ dysfunction in critically ill patients. Recent data suggests that early aggressive resuscitation of critically ill patients, which limits and/or reverses tissue dysoxia may prevent progression to organ dysfunction and improve outcome. The traditional clinical and laboratory markers used to assess tissue dysoxia are, however, insensitive and have numerous limitations. Regional carbon dioxide monitoring appears to be ideally suited to monitoring the adequacy of resuscitation. This review provides an update on this evolving technology.

## Recent findings

Gastric intramucosal carbon dioxide as measured by gastric tonometry has proven to be useful as a prognostic marker, in evaluating the response to specific therapeutic interventions and as an end point of resuscitation. Gastric tonometry is, however, cumbersome and has a number of limitations that may have prevented its widespread adoption. The measurement of carbon dioxide in the sublingual mucosa by sublingual capnometry is technically simple, noninvasive, and provides near instantaneous information. Clinical studies have demonstrated a good correlation between gastric intramucosal carbon dioxide and sublingual mucosa carbon dioxide. Sublingual mucosa carbon dioxide responds more rapidly to therapeutic interventions than does gastric intramucosal carbon dioxide and may be a better prognostic marker.

## Summary

Sublingual capnometry may be the ideal technology for guiding early goal directed therapy. This technology may be useful for monitoring tissue oxygenation, titrating therapeutic interventions, and as an end point for resuscitation in critically ill and injured patients.

## Keywords

critical care, gastric tonometry, sepsis, shock, sublingual capnometry, tissue carbon dioxide, tissue oxygenation

## Introduction

Despite improvements in resuscitation and supportive care, progressive organ dysfunction occurs in a large proportion of patients with acute, life-threatening illnesses. It has been proposed that the multi-organ dysfunction syndrome (MODS) of the critically ill is a consequence of tissue dysoxia due to inadequate oxygen delivery, often exacerbated by microcirculatory changes and increased tissue metabolic demands [1,2]. This may be further complicated by cytopathic hypoxia due to mitochondrial dysfunction [3,4]. A number of different but mutually compatible mechanisms might foster the development of cytopathic hypoxia. These mechanisms include the diminished delivery of pyruvate into the mitochondrial tricarboxylic acid (TCA) cycle, the inhibition of key mitochondrial enzymes that are involved in either the TCA cycle or the electron transport chain, and the activation of the enzyme poly(ADP-ribose) polymerase (PARP-1) [3–5].

In patients with sepsis tissue dysoxia may be an early finding. Experimental data suggests that tissue dysoxia occurring in the early phase of sepsis is predominantly due to decreased oxygen delivery, however, with time cytopathic hypoxia may play an increasing role [6]. Emerging data suggests that early aggressive resuscitation of critically ill patients may limit and/or reverse tissue dysoxia, progression to organ, and improve outcome. In a landmark study, Rivers *et al.* [7] demonstrated that a protocol of early goal directed therapy, reduces organ failure and improves survival. Similarly, Levy *et al.* [8] have demonstrated that the early (within 24 hours) reversal of shock, renal and respiratory failure was associated with improved survival in patients with sepsis. Nguyen *et al.* [9] have shown that the rate of clearance of lactate within 6 hours of presentation to the emergency room was predictive of outcome. Likewise, data from the ENHANCE study demonstrated that if activated protein C (Drotrecogin alpha-activated) is administered within 24 hours in patients with severe sepsis the mortality was 33% compared with 52% if the drug was administered after the second hospital day ( $P = 0.015$ ) [10]. These clinical studies suggest that early aggressive resuscitation that restores oxygen delivery and microcirculatory flow may limit tissue dysoxia, organ dysfunction, and ultimately patient outcome. This suggests that the end points of resuscitation should be indices of tissue oxygenation. However, the traditional clinical and laboratory markers used to assess tissue dysoxia are insensitive and have numerous limitations. Regional carbon dioxide ( $\text{CO}_2$ ) monitoring appears to be an evolving technology

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ideally suited to monitoring the adequacy of tissue oxygenation in the critically ill patient.

### **Assessing the adequacy of resuscitation**

The assessment of intravascular volume and the adequacy of volume resuscitation are amongst the most difficult clinical challenges. Systolic blood pressure, heart rate, and urine output change minimally in early shock. Furthermore, these variables provide little information as to the adequacy of tissue perfusion. Hypotension, tachycardia, cold extremities, decreased urine output, and poor capillary refill are only present in patients who have lost in excess of 30% of their blood volume (class III hemorrhage) [11]. The central venous pressure (CVP) and the changes in the CVP in response to volume loading are poor indicators of intravascular volume and recruitable cardiac index [12,13<sup>••</sup>]. In the study by Rivers *et al.* [7], the central venous oxygen saturation (ScvO<sub>2</sub>) was used as the end point of resuscitation in the intervention group, while in the control group treatment was guided by standard clinical end points including the CVP. While this study clearly demonstrates the value of early aggressive volume resuscitation, and while ScvO<sub>2</sub> does correlate quite well with SmvO<sub>2</sub>, the use of ScvO<sub>2</sub> to guide early resuscitation is not practical and has important limitations (see below) [14,15].

While flow to the brain and myocardium are preserved in patients with compensated shock, splanchnic and renal perfusion may be seriously compromised [16]. The gut is highly susceptible to diminished tissue perfusion and oxygenation as it has a higher critical oxygen requirement than the whole body and other vital organs, and the mucosal counter-current microcirculation renders the villi particularly vulnerable to ischemia [17,18]. Splanchnic hypoperfusion leads to both functional and structural changes in the gut mucosa with increased permeability and translocation of bacteria and bacterial products [17]. Increased mucosal permeability has been strongly associated with the development of the multi-organ dysfunction syndrome [17,19].

### **Global indices of tissue dysoxia**

While commonly measured in critically ill patients, blood lactate is an insensitive marker of tissue dysoxia [20–22]. If glycolysis occurs at a more rapid rate than is necessary for oxidative metabolism, some pyruvate may not be oxidatively metabolized in the Krebs cycle and will be converted to lactate. The result will be a concomitant increase in both pyruvate and lactate with an unchanged lactate/pyruvate ratio (L/P) [23]. Gore and co-workers measured lactate and pyruvate concentrations and the rates of pyruvate production and oxidation prior to and after dichloroacetate (DCA) administration in septic patients with severe lactic acidosis [5]. The patients in this report had significantly elevated levels of glucose, lac-

tate, and pyruvate (normal L/P ratio), with an increase in oxygen consumption and a significant decrease in glucose, lactate, and pyruvate (unchanged L/P ratio) after the administration of DCA.

It was concluded from this study that accumulation of lactate during sepsis was due to a markedly increased rate of pyruvate production. Furthermore, the arterial blood lactate level depends upon the rate of production as well as the rate of metabolism by the liver (Cori cycle). Due to decreased splanchnic blood flow and hepatocellular dysfunction, lactate removal may be impaired in critically ill patients.

Mixed venous oxygen saturation (SmvO<sub>2</sub>) measured in either the pulmonary artery (with a pulmonary artery catheter) or the right atrium (with a central venous catheter) has been used to monitor critically ill patients and their response to therapeutic interventions [7,15,24–26]. However, the SmvO<sub>2</sub> has been demonstrated to correlate poorly with cardiac index and indices of tissue dysoxia, particularly in patients with sepsis [21,27]. Patients with sepsis and liver disease may have a high SmvO<sub>2</sub> despite evidence of poor tissue perfusion [28,29]. This may be a consequence of the microcapillary shunting and/or the inability of the tissues to use oxygen (cytopathic hypoxia) [1,3,4,30].

### **Tissue CO<sub>2</sub> monitoring as an index of regional tissue oxygenation**

The development of tissue dysoxia in the gastrointestinal tract appears to be a common and early finding in patients with deranged hemodynamics. Dantzker [31] has suggested that the gastrointestinal tract may be the 'canary of the body', with gastrointestinal dysoxia an 'early warning of impending trouble.' Because direct gastrointestinal oxygen uptake and delivery are difficult to assess in patients, indirect techniques have been developed. Gastric tonometry and sublingual capnometry are based on the principle that tissue CO<sub>2</sub> levels rise sharply in conditions associated with poor tissue perfusion [27,28,32–36]. The increased tissue CO<sub>2</sub> (in part) is due to intracellular buffering of excesses of hydrogen ions by bicarbonate. The excesses of hydrogen ions are, in turn, traced to the degeneration of high-energy phosphate compounds during tissue dysoxia. Krebs *et al.* [37] noted that the acidosis that occurs during increased glycolysis caused by inadequate cellular oxygen delivery is produced by ATP that is continually being hydrolyzed.

The interpretation of tissue pCO<sub>2</sub> is however, relatively complex and determined by three variables, namely; the arterial CO<sub>2</sub> content (CaCO<sub>2</sub>), regional blood flow, and tissue CO<sub>2</sub> production (aerobic or anaerobic). In stable respiratory conditions when CaCO<sub>2</sub> is constant, tissue CO<sub>2</sub> essentially reflects the balance between tissue blood flow

and local CO<sub>2</sub> production. CO<sub>2</sub> production occurs in both aerobic and anaerobic conditions. Increases in aerobic metabolism are associated with higher CO<sub>2</sub> production by the cells, which is generally associated with parallel increase in blood flow, so that tissue PCO<sub>2</sub> does not increase (wash-out phenomenon). When oxygen delivery decreases and reaches a critical value, aerobiosis can no longer be sustained and aerobic production of CO<sub>2</sub> by cells increases.

It has been demonstrated that changes in gastrointestinal mucosal pCO<sub>2</sub> (PimCO<sub>2</sub>) mirrors changes in gastrointestinal oxygen uptake during progressive flow stagnation and that increases in PimCO<sub>2</sub> commence at the approximate onset of oxygen supply dependence [38,39]. Thus during low flow states, anaerobic CO<sub>2</sub> generation represents dysoxia, which can be detected by gut PimCO<sub>2</sub> measurement. It has been assumed that the increased tissue CO<sub>2</sub> seen in low flow states was a consequence of the intracellular buffering of excess hydrogen ions by bicarbonate. However, increased PimCO<sub>2</sub> may also result from flow stagnation and impaired washout of CO<sub>2</sub> produced aerobically or anaerobically [38,40–42]. Nevier *et al.* [43] reduced systemic oxygen delivery in a stepwise manner beyond critical oxygen delivery by lowering either the FiO<sub>2</sub> (hypoxic hypoxia) or reducing blood volume (ischemic hypoxia) in a pig model. While the ileal-arterial PCO<sub>2</sub> gap increased in both groups, the increase was much more dramatic in the ischemic group, with the pCO<sub>2</sub> only increasing at very low FiO<sub>2</sub> levels (0.08) in the hypoxic group. Furthermore, in the hypoxic group the arterial lactate increased much earlier and to a greater degree than the ileal-arterial pCO<sub>2</sub> gap. This study indicates that the increased tissue CO<sub>2</sub> noted in low flow states is predominantly due to flow stagnation. This suggests that tissue CO<sub>2</sub> monitoring may be useful in low flow states (compensated or uncompensated shock) while lactate levels may be more useful in hypoxic or anemic hypoxia. From a practical point, almost all clinical states of decreased oxygen delivery (tissue hysoxia) result from decreased tissue perfusion and flow stagnation rather than reduced oxygen carrying capacity (which is in itself very easy to diagnose). As such the increased tissue CO<sub>2</sub> which occurs in critically ill patients is likely to be mainly due to flow stagnation with smaller components due anaerobic CO<sub>2</sub> production (due to decreased oxygen delivery and cytopathic hypoxia).

### Gastric tonometry

Tonometry is simply a method used to equilibrate gas tension between two compartments. Either gas (e.g., air) or fluid (e.g., normal saline) can be used as the solvent for equilibration of the gas in question (i.e., carbon dioxide). A gastric tonometer is a standard nasogastric tube with a silicone balloon at its distal end. With conventional saline tonometry, the balloon of the tonometer is filled with normal saline and allowed to equilibrate (ideally for 90 minutes). Once equilibrated, the balloon is aspirated

and the pCO<sub>2</sub> of the fluid determined using a standard blood gas analyzer. Air tonometry, particularly in conjunction with an automated system has a number of advantages when compared with conventional saline tonometry. Unlike saline tonometry, in-vitro studies have demonstrated that air tonometry has a very low bias with excellent precision [44–46]. Furthermore, automated air tonometry has an equilibration time of less than 20 minutes and being fully automated may eliminate potential sources of error associated with saline tonometry.

Monitoring of gastric intramucosal pH (pHi) was initially used to assess splanchnic dysoxia. However the calculation of the pHi involves a number of assumptions (serum HCO<sub>3</sub><sup>-</sup> equal to splanchnic HCO<sub>3</sub><sup>-</sup>) that may be incorrect. Consequently, the PCO<sub>2</sub> gap, defined as the difference between the gastric mucosal PCO<sub>2</sub> (PgCO<sub>2</sub>) and the arterial PCO<sub>2</sub> may more specifically reflect the adequacy of gastric mucosal blood flow, and is not influenced by systemic acid-base status [47].

Gastric intramucosal acidosis (pHi) and intramucosal hypercarbia (PCO<sub>2</sub> gap) have been demonstrated to be a marker of gastric mucosal dysoxia and a predictor of morbidity and mortality in critically ill patients [22,33,39,47–55]. Levy *et al.* [56] measured gastric mucosal PCO<sub>2</sub> (by automated air-tonometry) in 95 consecutive critically ill patients on admission to the ICU and at 24 hours. By univariate analysis the pHi was significantly higher on admission and at 24 hours in the survivors as compared with the non-survivors. By multivariate analysis the organ failure score and the PCO<sub>2</sub> gap at 24 hours were independent predictors of outcome. The 28-day survival was 75% in patients with a PCO<sub>2</sub> gap of less than 20 mm Hg at 24 hours compared with a 28-day survival of 40% in those patients with a PCO<sub>2</sub> gap of greater than 20 mm Hg at 24 hours. This study supports the observation that the failure to improve organ function and tissue perfusion within 24 hours of admission to the ICU is associated with a poor outcome.

Gutierrez *et al.* [54] randomized critically ill ICU patients to a standard treatment group or a protocol group in which treatment was titrated to maintain the gastric intramucosal pH (pHi) greater than 7.35. Survival was significantly improved in the protocol sub-group whose initial pHi was greater than 7.35. This study provides further support to the argument that the early detection and treatment of tissue dysoxia may improve the outcome of critically ill patients. Once the ‘golden hours’ of resuscitation have lapsed and progressive tissue dysoxia has developed measures that improve tissue dysoxia are unlikely to improve outcome.

Silva *et al.* [57] reported the changes in systemic hemodynamics variables and PCO<sub>2</sub> gap in septic patients following

a fluid challenge. While the fluid challenge was associated with an increase in cardiac index, global measures of tissue oxygenation (S<sub>mv</sub>O<sub>2</sub> and oxygen consumption) were unchanged; there was, however, a significant fall in the PCO<sub>2</sub> gap. Furthermore, neither the baseline hemodynamic variables nor the change in cardiac index were related to the change in the change in the PCO<sub>2</sub> gap. The change in the PCO<sub>2</sub> gap was, however, highly associated with the baseline PCO<sub>2</sub> gap. These results provide further support that regional indices of tissue oxygenation (PCO<sub>2</sub> gap in this case) rather than global parameters should be used when initiating and titrating resuscitative measures in the critically ill.

The base excess (BE) has become the standard end point of resuscitation in trauma patients. Remarkably, while the BE has been demonstrated to be of prognostic value, it has never been assessed prospectively in trauma patients [58–64]. The use of the BE is based on the principle that tissue hypoxia associated with poor perfusion will result in the generation of hydrogen ions and a metabolic acidosis. However, it is likely that tissue hypoperfusion may occur in the absence of a significant change in the BE. Furthermore, as it requires time for the liver and kidney to regenerate bicarbonate [65], it can be expected that there will be a long lag phase between the correction of intravascular volume and normalization of the BE. In a rat hemorrhage model, Totapally *et al.* [66] demonstrated that the BE responded slowly to changes in intravascular volume and that there was a significant increase in the BE only when the mean arterial blood pressure fell by greater than 50%, however, changes in the esophageal CO<sub>2</sub>-gap closely mirrored changes in intravascular volume.

Kirton *et al.* have demonstrated that gastric intramucosal pH (pHi) correlates well with the degree of injury and that optimizing the pHi in the first 24 hours following trauma is associated with a reduction in the incidence of organ failure and death [67,68].

Ivatury *et al.* [53] randomized trauma patients to a protocol in which global (oxygen delivery index) or regional (gastric tonometry) indexes were used as the end points of resuscitation. Of the 44 patients with a pHi greater than 7.3 at 24 hours only 3 died (6.8%) as compared with 7 of 13 (53.9%) in whom the pHi was not optimized. This study is in keeping with the sepsis studies cited above, which suggest that the early correction of tissue hypoperfusion may improve outcome.

Gastric tonometry has been shown to be useful in titrating vasopressor support and determining which vasopressor agent or vasoactive drug combination improves gastric perfusion in critically ill patients. Several studies have demonstrated that dobutamine, dobutamine/norepinephrine combination, norepinephrine alone, or dopexamine will

increase gastric pHi or decrease PCO<sub>2</sub> gap compared with other agents or placebo in patients with sepsis or septic shock or high-risk surgical patients [69–74]. Paradoxically dopamine may cause gastric mucosal ischemia [69,70,75]. In a porcine shock model, treatment with low-dose dopamine hastened the development of gut ischemia [76]. It is postulated that dopamine causes precapillary vasoconstriction with diversion of blood flow away from the gut mucosa. ‘Low dose’ vasopressin has recently become popular in the management of septic shock. It has been suggested that sepsis is a vasopressin deficient state and that low doses of vasopressin may restore vascular tone without causing splanchnic ischemia [77–79]. However, van Haren *et al.* [80] have demonstrated that in norepinephrine dependent patients in septic shock, continuous infusions of low-dose vasopressin result in a significant increase in the PCO<sub>2</sub> gap compatible with splanchnic hypoperfusion.

In addition to the indications cited above, gastric tonometry has been demonstrated to be useful in predicting weaning outcome from mechanical ventilation [81–84]. These studies have demonstrated that patients who could not be weaned from mechanical ventilation or failed extubation had a significant fall in pHi during the weaning trial and/or a lower baseline pHi. It was postulated that the increased work of breathing during the weaning trial caused blood flow to be diverted away from the gut leading to splanchnic ischemia. Bocquillon *et al.* [81] demonstrated an increase in cardiac index and a decrease in gastric mucosal blood flow in patients who failed weaning. In this study the fall in the pHi was associated with an increase in arterial CO<sub>2</sub> rather than an increase in the gastric intramucosal-arterial PCO<sub>2</sub> gap.

### **Sublingual capnometry**

While air tonometry has simplified the measurement process (over saline tonometry) and eliminated the possible errors associated with the use of non-buffered saline this technology has a number of limitations [44–46,85,86]. Gastric tonometry is logistically and practically difficult and this may be the main factor that has prevented the widespread use of this technology. Gastric tonometry requires placement of a specialized naso-gastric tube. This precludes its use in pediatric patients and limits its use as a screening tool in patients who may not require nasogastric intubation. Equilibration of carbon dioxide between the gastric mucosa and the balloon is time dependent (about 15 minutes with air tonometry) and likely to be slowly responsive to therapeutic interventions. This slow equilibration time limits the use of gastric tonometry in the acute phase of resuscitation. Histamine type-2 blockers (H<sub>2</sub>-blockers) are routinely required to limit the intraluminal generation of carbon dioxide from gastric acid. Furthermore, enteral nutrition must be stopped at least 2 hours prior to each measurement; this is likely to interfere with the provision of nutritional support.

To overcome the potential limitations of gastric tonometry, Jin *et al.* [87] postulated that the very proximal gastrointestinal tract, namely, the tongue and/or sublingual mucosa, may serve as appropriate site for measurement of tissue PCO<sub>2</sub>. Although the tongue receives its blood supply from the internal carotid artery, the tongue may act functionally as part of the 'splanchnic circulation'. Indeed, Weil *et al.* [87] have demonstrated that with decreased perfusion pressure blood flow to the tongue and splanchnic bed fall to a similar degree. These authors have demonstrated an increase in sublingual PCO<sub>2</sub> (PslCO<sub>2</sub>) that was closely related to decreases in arterial pressure and cardiac index during circulatory shock produced by hemorrhage and sepsis, [35,88–90]. Furthermore, the increase in PslCO<sub>2</sub> closely tracked the increase in PimCO<sub>2</sub>. Similarly, Marik [27] has demonstrated an excellent correlation between the PimCO<sub>2</sub> and PslCO<sub>2</sub> ( $r = 0.78$ ;  $P < 0.001$ ) in a heterogeneous group of ICU patients. De Backer *et al.* [91] used orthogonal polarization spectral imaging to investigate the sublingual microcirculation in healthy controls and patients with severe sepsis. The proportion of perfused small vessels ( $< 20 \mu\text{m}$ ) was significantly reduced in patients with sepsis as compared with controls (48% vs 90%,  $P < 0.001$ ). In a follow-up study these authors have demonstrated an improvement in the sublingual microcirculation with an increase in perfused vessels density and an increase in the percentage of small perfused vessels with volume and pressor resuscitation [29].

The currently available system for measuring sublingual pCO<sub>2</sub> consists of a disposable pCO<sub>2</sub> sensor (which is placed under the tongue) and a battery-powered handheld instrument (CapnoProbe N80, Nellcor, CA). This technology is based on a carbon dioxide sensing optode containing a fluorescent indicator, which is excited by light conducted through an optical fiber, which then transmits the fluorescent emission back to the instrument.

Marik and Bankov [28] have shown that in critically ill ICU patients, the PslCO<sub>2</sub> better differentiates between survivors and non-survivors than lactate or S<sub>MV</sub>O<sub>2</sub> and is more responsive to therapy than either of these markers. These results were confirmed by Creteur *et al.* [29], who, in addition, demonstrated that the PslCO<sub>2</sub> correlated well with microvascular perfusion and that it was more responsive to therapy than the PimCO<sub>2</sub>. In patients with penetrating trauma, Baron *et al.* [92] demonstrated that the PslCO<sub>2</sub> correlated well with the degree of blood loss and was a good predictor of outcome.

## Conclusion

Sublingual capnometry is a technically simple, noninvasive, inexpensive technology that provides near instantaneous information as to the adequacy of tissue perfusion in critically ill and injured patients. This technology is

ideally suited to the emergency room, operating room, and ICU and PslCO<sub>2</sub> monitoring may prove to be a useful end point for goal directed resuscitation. The fact that sublingual capnometry does not require premedication with acid suppressive therapy nor discontinuation of enteral feeding is a major advantage over gastric tonometry. The clinical experience with sublingual capnometry is however limited, and additional studies are needed, which demonstrate the clinical utility of PslCO<sub>2</sub> monitoring and further refinements in the technology are required to allow continuous as well as intermittent measurements.

## References and recommended reading

Papers of particular interest, published within the annual period of review, have been highlighted as:

- of special interest
- of outstanding interest

- 1 Ince C, Sinaasappel M. Microcirculatory oxygenation and shunting in sepsis and shock. *Crit Care Med* 1999; 27:1369–1377.
- 2 Beal AL, Cerra FB. Multiple organ failure syndrome in the 1990's: systemic inflammatory response and organ dysfunction. *JAMA* 1994; 271:226–233.
- 3 Fink MP. Bench-to-bedside review: cytopathic hypoxia. *Crit Care* 2002; 6: 491–499.
- 4 Fink MP. Cytopathic hypoxia. Is oxygen use impaired in sepsis as a result of an acquired intrinsic derangement in cellular respiration? *Crit Care Clin* 2002; 18:165–175.
- 5 Gore DC, Jahoor F, Hibbert JM, *et al.* Lactic acidosis during sepsis is related to increased pyruvate production, not deficits in tissue oxygen availability. *Ann Surg* 1996; 224:97–102.
- 6 Simonson SG, Welty-Wolf K, Huang YC, *et al.* Altered mitochondrial redox responses in gram negative septic shock in primates. *Circ Shock* 1994; 43:34–43.
- 7 Rivers E, Nguyen B, Havstad S, *et al.* Early goal-directed therapy in the treatment of severe sepsis and septic shock. *N Engl J Med* 2001; 345:1368–1377.
- 8 Levy MM, Macias WL, Russel JA, *et al.* Failure to improve during the first day of therapy is predictive of 28-day mortality in severe sepsis [abstract]. *Chest* 2003; 124:120S.
- 9 Nguyen HB, Rivers EP, Knoblich BP, *et al.* Early lactate clearance is associated with improved outcome in severe sepsis and septic shock. *Crit Care Med* 2004; 32:1637–1642.
- 10 Bernard GR, Margolis BD, Shanies HM, *et al.* Extended evaluation of recombinant human activated protein C United States Trial (ENHANCE US): a single-arm, phase 3B, multicenter study of drotrecogin alfa (activated) in severe sepsis. *Chest* 2004; 125:2206–2216.
- 11 Shock: Advanced Trauma Life Support for Doctors; Student Course Manual. 6th ed. Chicago, IL: American College of Surgeons; 1997: 87–112.
- 12 Michard F, Teboul JL. Predicting fluid responsiveness in ICU patients: a critical analysis of the evidence. *Chest* 2002; 121:2000–2008.
- 13 Kumar A, Anel R, Bunnell E, *et al.* Pulmonary artery occlusion pressure and central venous pressure fail to predict ventricular filling volume, cardiac performance, or the response to volume infusion in normal subjects. *Crit Care Med* 2004; 32:691–699.
- This is an elegant study that demonstrates that many of the standard pressure derived variables used to predict volume responsiveness are inaccurate.
- 14 Marik PE, Varon J. Goal-directed therapy in sepsis. *N Engl J Med* 2002; 346:1025.
- 15 Reinhart K, Kuhn HJ, Hartog C, *et al.* Continuous central venous and pulmonary artery oxygen saturation monitoring in the critically ill. *Intensive Care Med* 2004; 30:1572–1578.
- 16 Ba ZF, Wang P, Koo DJ, *et al.* Alterations in tissue oxygen consumption and extraction after trauma and hemorrhagic shock. *Crit Care Med* 2000; 28: 2837–2842.
- 17 Pastores SM, Katz DP, Kvetan V. Splanchnic ischemia and gut mucosal injury in sepsis and the multiple organ dysfunction syndrome. *Am J Gastroenterol* 1996; 91:1697–1710.

- 18 Nelson D, Beyer C, Samsel R, *et al.* Pathologic supply dependence of systemic and intestinal O<sub>2</sub> uptake during bacteremia in the dog. *J Appl Physiol* 1987; 63:1487–1489.
- 19 Doig CJ, Sutherland LR, Sandham JS, *et al.* Increased intestinal permeability is associated with the development of multiple organ dysfunction syndrome in critically ill ICU patients. *Am J Respir Crit Care Med* 1998; 158:444–451.
- 20 Hotchkiss RS, Karl IE. Reevaluation of the role of cellular hypoxia and bioenergetics failure in sepsis. *JAMA* 1992; 267:1503–1510.
- 21 Marik PE, Varon J. The hemodynamic derangements in sepsis: Implications for treatment strategies. *Chest* 1998; 114:854–860.
- 22 Marik PE. Gastric intramucosal pH. A better predictor of multiorgan dysfunction syndrome and death than oxygen-derived variables in patients with sepsis. *Chest* 1993; 104:225–229.
- 23 Levy B, Bollaert PE, Charpentier C, *et al.* Comparison of norepinephrine and dobutamine to epinephrine for hemodynamics, lactate metabolism, and gastric tonometric variables in septic shock: a prospective, randomized, study. *Intensive Care Med* 1997; 23:282–287.
- 24 Astiz ME, Rackow EC, Kaufman B, *et al.* Relationship of oxygen delivery and mixed venous oxygenation to lactic acidosis in patients with sepsis and acute myocardial infarction. *Crit Care Med* 1988; 16:655–658.
- 25 Mahutte CK, Jaffe MB, Sasse SA, *et al.* Relationship of thermodilution cardiac output to metabolic measurements and mixed venous oxygen saturation. *Chest* 1993; 104:1236–1242.
- 26 Vaughn S, Puri VK. Cardiac output changes and continuous mixed venous oxygen saturation measurement in the critically ill. *Crit Care Med* 1988; 16: 495–498.
- 27 Marik PE. Sublingual capnography: a clinical validation study. *Chest* 2001; 120:923–927.
- 28 Marik PE, Bankov A. Sublingual capnometry versus traditional markers of tissue oxygenation in critically ill patients. *Crit Care Med* 2003; 31:818–822.
- 29 Creteur J, De Backer D, Sakr Y, *et al.* Sublingual PCO<sub>2</sub> monitoring in patients with septic shock [abstract]. *Crit Care Med* 2003; 30:A19.
- 30 Ince C. The microcirculation unveiled. *Am J Respir Crit Care Med* 2002; 166:1–4.
- 31 Dantzer DR. The gastrointestinal tract. The canary of the body? *JAMA* 1993; 270:1247–1248.
- 32 Fink MP. Tissue capnometry as a monitoring strategy for critically ill patients: just about ready for prime time. *Chest* 1998; 114:667–670.
- 33 Gutierrez G, Brown SD. Gastrointestinal tonometry: a monitor of regional dysoxia. *New Horiz* 1996; 4:413–419.
- 34 Sato Y, Weil MH, Tang W. Tissue hypercarbic acidosis as a marker of acute circulatory failure (shock). *Chest* 1998; 114:263–274.
- 35 Weil MH, Nakagawa Y, Tang W, *et al.* Sublingual capnometry: a new noninvasive measurement for diagnosis and quantitation of severity of circulatory shock. *Crit Care Med* 1999; 27:1225–1229.
- 36 Marik P. Gastric tonometry: the canary sings once again. *Crit Care Med* 1998; 26:809–810.
- 37 Krebs HA, Woods HF, Alberti KGMM. Hyperlactataemia and lactic acidosis. *Essays Biochem* 1970; 1:81–103.
- 38 Grum CM, Fiddian-Green RG, Pittenger GL, *et al.* Adequacy of tissue oxygenation in intact dog intestine. *J Appl Physiol* 1984; 56:1065–1069.
- 39 Schlichtig R, Bowles SA. Distinguishing between aerobic and anaerobic appearance of dissolved CO<sub>2</sub> in intestine during low flow. *J Appl Physiol* 1994; 76:2443–2451.
- 40 Guzman JA, Kruse JA. Development and validation of a technique for continuous monitoring of gastric intramucosal pH. *Am J Respir Crit Care Med* 1996; 153:694–700.
- 41 Salzman AL, Wang H, Wollert PS, *et al.* Endotoxin-induced ileal mucosal hyperpermeability in pigs: role of tissue acidosis. *Am J Physiol* 1994; 266: G633–G646.
- 42 Dubin A, Murias G, Estenssoro E, *et al.* Intramucosal-arterial PCO<sub>2</sub> gap fails to reflect intestinal dysoxia in hypoxic hypoxia. *Crit Care* 2002; 6:514–520.
- 43 Neviere R, Chagnon JL, Teboul JL, *et al.* Small intestine intramucosal PCO<sub>2</sub> and microvascular blood flow during hypoxic and ischemic hypoxia. *Crit Care Med* 2002; 30:379–384.
- 44 Graf J, Konigs B, Mottaghy K, *et al.* In vitro validation of gastric air tonometry using perfluorocarbon FC 43 and 0.9% sodium chloride. *Br J Anaesth* 2000; 84:497–499.
- 45 Barry B, Mallick A, Hartley G, *et al.* Comparison of air tonometry with gastric tonometry using saline and other equilibrating fluids: an in vivo and in vitro study. *Intensive Care Med* 1998; 24:777–784.
- 46 Tzelepis G, Kadas V, Michalopoulos A, *et al.* Comparison of gastric air tonometry with standard saline tonometry. *Intensive Care Med* 1996; 22:1239–1243.
- 47 Schlichtig R, Mehta N, Gayowski TJP. Tissue-arterial PCO<sub>2</sub> difference is a better marker of ischemia than intramucosal pH (pHi) or arterial pH-pHi difference. *J Crit Care* 1996; 11:51–56.
- 48 Antonsson JB, Boyle CC, Kruthoff KL, *et al.* Validation of tonometric measurement of gut intramucosal pH during endotoxemia and mesenteric occlusion in pigs. *Am J Physiol* 1990; 259:G519–G523.
- 49 Montgomery AM, Hartmann K, Jonsson K, *et al.* Intramucosal pH measurement with tonometers for detecting gastrointestinal ischemia in porcine hemorrhagic shock. *Circ Shock* 1989; 29:319–327.
- 50 Elizalde JI, Hernández C, Lach J, *et al.* Gastric intramucosal acidosis in mechanically ventilated patients: Role of mucosal blood flow. *Crit Care Med* 1998; 26:827–833.
- 51 Maynard N, Bihari D, Beale R, *et al.* Assessment of splanchnic oxygenation by gastric tonometry in patients with acute circulatory failure. *JAMA* 1993; 270:1203–1210.
- 52 Friedman G, Berlot G, Kahn RJ, *et al.* Combined measurements of blood lactate concentrations and gastric intramucosal pH in patients with severe sepsis. *Crit Care Med* 1995; 23:1184–1193.
- 53 Ivatury RR, Simon RJ, Islam S, *et al.* A prospective randomized study of end points of resuscitation after major trauma: global oxygen transport indices versus organ-specific gastric mucosal pH. *J Am Coll Surg* 1996; 183:145–154.
- 54 Gutierrez G, Palizas F, Doglio G, *et al.* Gastric intramucosal pH as a therapeutic index of tissue oxygenation in critically ill patients. *Lancet* 1992; 339:195–199.
- 55 Mythen MG, Webb AR. Perioperative plasma volume expansion reduces the incidence of gut mucosal hypoperfusion during cardiac surgery. *Arch Surg* 1995; 130:423–429.
- 56 Levy B, Gawalkiewicz P, Vallet B, *et al.* Gastric capnometry with air-automated tonometry predicts outcome in critically ill patients. *Crit Care Med* 2003; 31:474–480.
- 57 Silva E, De Backer D, Creteur J, *et al.* Effects of fluid challenge on gastric mucosal PCO<sub>2</sub> in septic patients. *Intensive Care Med* 2004; 30:423–429.
- 58 Siegel JH, Rivkind AI, Dalal S, *et al.* Early physiologic predictors of injury severity and death in blunt multiple trauma. *Arch Surg* 1990; 125:498–508.
- 59 Davis JW, Parks SN, Kaups KL, *et al.* Admission base deficit predicts transfusion requirements and risk of complications. *J Trauma* 1996; 41:769–774.
- 60 Rutherford EJ, Morris JA Jr, Reed GW, *et al.* Base deficit stratifies mortality and determines therapy. *J Trauma* 1992; 33:417–423.
- 61 Kincaid EH, Chang MC, Letton RW, *et al.* Admission base deficit in pediatric trauma: a study using the National Trauma Data Bank. *J Trauma* 2001; 51:332–335.
- 62 Porter JM, Ivatury RR. In search of the optimal end points of resuscitation in trauma patients: a review. *J Trauma* 1998; 44:908–914.
- 63 Ivatury RR, Sugerman H. In quest of optimal resuscitation: tissue specific, on to the microcirculation. *Crit Care Med* 2000; 28:3102–3103.
- 64 Rixen D, Raum M, Bouillon B, *et al.* Base deficit development and its prognostic significance in posttrauma critical illness: an analysis by the trauma registry of the Deutsche Gesellschaft für Unfallchirurgie. *Shock* 2001; 15:83–89.
- 65 Stacpoole PW. Lactic acidosis. *Endocrinol Metab Clin North Am* 1993; 22:221–245.
- 66 Totapally BR, Fakioglu H, Torbati D, *et al.* Esophageal capnometry during hemorrhagic shock and after resuscitation in rats. *Crit Care* 2003; 7:79–84.
- 67 Kirton OC, Windsor J, Wedderburn R, *et al.* Failure of splanchnic resuscitation in the acutely injured trauma patient correlates with multiple organ system failure and length of stay in the ICU. *Chest* 1998; 113:1064–1069.
- 68 Barquist E, Kirton O, Windsor J, *et al.* The impact of antioxidant and splanchnic-directed therapy on persistent uncorrected gastric mucosal pH in the critically injured trauma patient. *J Trauma* 1998; 44:355–360.
- 69 Marik PE, Mohedin M. The contrasting effects of dopamine and norepinephrine on systemic and splanchnic oxygen utilization in hyperdynamic sepsis. *JAMA* 1994; 272:1354–1357.
- 70 Neviere R, Mathieu D, Chagnon JL, *et al.* The contrasting effects of dobutamine and dopamine on gastric mucosal perfusion in septic patients. *Am J Respir Crit Care Med* 1996; 154:1684–1688.
- 71 Neviere R, Chagnon JL, Vallet B, *et al.* Dobutamine improves gastrointestinal mucosal blood flow in a porcine model of endotoxic shock. *Crit Care Med* 1997; 25:1371–1377.

- 72 Duranteau J, Sitbon P, Teboul JL, *et al.* Effects of epinephrine, norepinephrine, or the combination of norepinephrine and dobutamine on gastric mucosa in septic shock. *Crit Care Med* 1999; 27:893–900.
- 73 Meier-Hellman A, Bredle D, Specht M, *et al.* Dopexamine increases splanchnic blood flow but decreases gastric mucosal pH in severe septic patients treated with dobutamine. *Crit Care Med* 1999; 27:2166–2177.
- 74 Steinberg S, Azar G, Love R, *et al.* Dopexamine prevents depression of mesenteric blood flow caused by positive end-expiratory pressure in rats. *Surgery* 1996; 120:597–601.
- 75 Olson D, Pohlman A, Hall JB. Administration of low-dose dopamine to non-oliguric patients with sepsis syndrome does not raise intramucosal gastric pH nor improve creatinine clearance. *Am J Respir Crit Care Med* 1996; 154:1664–1670.
- 76 Segal JM, Phang PT, Walley KR. Low-dose dopamine hastens onset of gut ischemia in a porcine model of hemorrhagic shock. *J Appl Physiol* 1992; 73:1159–1164.
- 77 Landry DW, Levin HR, Gallant EM, *et al.* Vasopressin pressor hypersensitivity in vasodilatory septic shock. *Crit Care Med* 1997; 25:1279–1282.
- 78 Law AW, Gales MA. Octreotide or vasopressin for bleeding esophageal varices. *Ann Pharmacother* 1997; 31:237–238.
- 79 Raid IA. Role of vasopressin deficiency in the vasodilation of septic shock. *Circulation* 1997; 95:1108–1110.
- 80 van Haren FM, Rozendaal FW, van der Hoeven JG. The effect of vasopressin on gastric perfusion in catecholamine-dependent patients in septic shock. *Chest* 2003; 124:2256–2260.
- 81 Bocquillon N, Mathieu D, Neviere R, *et al.* Gastric mucosal pH and blood flow during weaning from mechanical ventilation in patients with chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 1999; 160:1555–1561.
- 82 Hurtado FJ, Beron M, Olivera W, *et al.* Gastric intramucosal pH and intraluminal PCO<sub>2</sub> during weaning from mechanical ventilation. *Crit Care Med* 2001; 29:70–76.
- 83 Bouachour G, Guiraud MP, Gouello JP, *et al.* Gastric intramucosal pH: an indicator of weaning outcome from mechanical ventilation in COPD patients. *Eur Respir J* 1996; 9:1868–1873.
- 84 Mohsenifar Z, Hay A, Hay J, *et al.* Gastric intramural pH as a predictor of success or failure in weaning patients from mechanical ventilation. *Ann Intern Med* 1993; 119:794–798.
- 85 Taylor DE, Gutierrez G, Clark C, *et al.* Measurement of gastric mucosal carbon dioxide tension by saline and air tonometry. *J Crit Care* 1997; 12:208–213.
- 86 Dullenkopf A, Cornelius A, Gerber AC, *et al.* Factors affecting performance of air tonometry using the TONOCAP. *Anaesth Intensive Care* 2002; 30:794–799.
- 87 Jin X, Weil MH, Sun S, *et al.* Decreases in organ blood flows associated with increases in sublingual PCO<sub>2</sub> during hemorrhagic shock. *J Appl Physiol* 1998; 85:2360–2364.
- 88 Pernat A, Weil MH, Tang W, *et al.* Effects of hyper- and hypoventilation on gastric and sublingual PCO<sub>2</sub>. *J Appl Physiol* 1999; 87:933–937.
- 89 Nakagawa Y, Weil MH, Tang W, *et al.* Sublingual capnometry for diagnosis and quantitation of circulatory shock. *Am J Respir Crit Care Med* 1998; 157:1838–1843.
- 90 Povoas HP, Weil MH, Tang W, *et al.* Comparisons between sublingual and gastric tonometry during hemorrhagic shock. *Chest* 2000; 118:1127–1132.
- 91 De Backer D, Creteur J, Preiser JC, *et al.* Microvascular blood flow is altered in patients with sepsis. *Am J Respir Crit Care Med* 2002; 166:98–104.
- 92 Baron BJ, Inerrt R, Zehtabchi S, *et al.* Diagnostic utility of sublingual PCO<sub>2</sub> for detecting hemorrhage in patients with penetrating trauma [abstract]. *Acad Emerg Med* 2002; 9:492.